

*Perspective*

## Immunometabolism and Covid-19: Could Lifelong Exercise Training Have a Protective Effect?

Luciele Guerra Minuzzi<sup>1,2</sup>, Ana Maria Teixeira<sup>2</sup>,  
Ronaldo V. Thomatieli-Santos<sup>3</sup>, José C. Rosa-Neto<sup>4</sup>, Fábio S. Lira<sup>1,\*</sup>

<sup>1</sup> Exercise and Immunometabolism Research Group, Department of Physical Education, Universidade Estadual Paulista “Julio de Mesquita Filho”, Presidente Prudente, São Paulo 19060-900, Brazil

<sup>2</sup> Research Center for Sport and Physical Activity, Faculty of Sports Science and Physical Education, University of Coimbra, Coimbra 3040-248, Portugal

<sup>3</sup> Department of Bioscience, Universidade Federal de São Paulo, Santos 11015-020, Brazil

<sup>4</sup> Institute of Biomedical Sciences, Universidade de São Paulo, São Paulo 05508-000, Brazil

\* Correspondence: Fábio S. Lira, Email: Fabio.lira@unesp.br;  
Tel.: +55-18-3229-5906.

---

### ABSTRACT

The world is experiencing a severe new pandemic, in which the elderly afflicted with chronic diseases are the most affected. The aging of the immune system (immunosenescence) reduces its efficacy against viral infections and increases its susceptibility to repeated acute infections, such as the flu. The improvement of the immune system functioning leading to a reduced incidence of infections can be achieved with regular physical exercise, besides its countless other benefits. The immunosenescence delay in master athletes, protecting them from possible viral infections, has been recently shown. Here the role of aerobic exercise training as an immune system fine-tuning regulator was discussed, focusing on lifelong athletes and specifically on the age-impaired antibody production in immunized elderly and the effects of lifelong physical exercise on the anti-inflammatory and vaccine response optimization. Moreover, the aerobic training effects on the natural killer (NK) cell activity and the underlying mechanisms responsible for a better antiviral response in active elderly and/or master athletes were addressed. It was hypothesized that lifelong exercise training delays age-related decrements in immunity by remodeling the metabolism of different cells (e.g., NK cells), creating a metabolic scenario that in turn improves the immune system's viral response. Lifelong exercisers present a preserved immune response to exercise, indicating that they are better prepared to respond to new immune challenges. Thus, master athletes and lifelong exercisers are possibly protected against or could mitigate the COVID-19 disease.

### Open Access

Received: 13 August 2020

Accepted: 30 October 2020

Published: 27 November 2020

Copyright © 2020 by the author(s). Licensee Hapres, London, United Kingdom. This is an open access article distributed under the terms and conditions of [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/).

**KEYWORDS:** exercise training; master athletes; viral infections; NK cells; COVID-19

---

## INTRODUCTION

Viral infections are very common. Indeed, all individuals have already experienced symptoms, such as sore throat, fever, and headaches, among others. Typically, it takes up to 7 days for the immune system to resolve viral infections. Since birth, humans are exposed to different types of viruses, leading the human body to quickly trigger an immune response to eliminate the virus, incorporating a recognition pattern (innate immunity), and producing memory cells (acquired immunity). Thus, the immune system will act more efficiently if the virus is again found in the circulation.

Regular physical exercise is closely related to the immune response's response to viral infections. Studies have shown the enhanced role of the immune system and the enhanced anti-inflammatory response to chronic and acute exercise sessions of different intensities in subjects who regularly exercise and/or lifelong athletes compared to sedentary subjects [1–7].

The intensity and duration of physical exercise are crucial factors to determine the different alterations in the immune system. Acute physical exercise, when performed, can generate several transient changes in the body. Besides, physiological adaptations are also developed in response to stress to reestablish homeostasis, greatly influencing the immune response [8–10]. Throughout the athlete's life, immune cells undoubtedly suffer from metabolism transformation when adapting to periods of energy overload and physiological stress, thus creating a metabolic landscape that, in turn, improves immunosurveillance and viral response. This metabolic reprogramming of immune cells may be particularly important for natural killer (NK) cells. NK cells are the immune cells most responsive to exercise, acutely mobilized into the circulation, during exercise performance [11,12]. Over time, these transient exercise-induced alterations promote an increase in selective lymphocyte subsets, enhanced immunosurveillance, and lower inflammation [13], which may be of particular clinical value for infected individuals. Furthermore, physical exercise up-regulates NK cell proliferation while stimulating their maturation and activation [14]. The exercise-dependent metabolic reprogramming of NK cells remains still unexplored. However, it is widely recognized that chronic training-induced alterations include improved metabolic health. Herein, it is hypothesized that lifelong exercise training induces metabolism remodeling of different cells, particularly NK cells, creating a metabolic landscape that improves viral response.

The world has been greatly affected by the COVID-19 pandemic. The pathogenic mechanism of SARS-CoV-2 infection is complex and still poorly understood. SARS-CoV-2 can cross mucous membranes, especially the

nasal and laryngeal mucosa, entering the lungs through the respiratory tract. As a result, SARS-CoV-2 can migrate, attacking target organs expressing the angiotensin-converting enzyme 2, such as the lungs, heart, kidney system, gastrointestinal tract, and endothelial cells [15–18]. After a few days of regression of signs and symptoms, more precisely between the 7th and 14th days after the onset of infection, SARS-CoV-2 starts a second attack. The worsening of clinical conditions in this second phase may be due to the acquired immunity's inability to respond to the challenges imposed by SARS-CoV-2. During the second phase, a reduction in B lymphocytes can occur, affecting the antibody production in patients [19]. The total number of lymphocytes and CD4<sup>+</sup> T, CD8<sup>+</sup> T, B, and NK cells are decreased in COVID-19 patients, with a lymphopenia of about 70% [20].

In contrast, lifelong trained older adults (presenting an improved physical fitness condition) show preserved CD4<sup>+</sup> naive T-cell population and decreased senescent T-cell percentages [1]. It is possible that maintaining high levels of aerobic fitness during the natural course of aging may help to increase immunosurveillance by preserving the number of CD4<sup>+</sup> naive T cells [1], which in turn could be critical for detecting SARS-CoV-2-infected cells.

Therefore, in this perspective article, the role of aerobic exercise training as a fine-tuning regulator of the immune system was addressed, focusing on lifelong athletes. The effects of lifelong physical exercise on improving the anti-inflammatory and vaccine responses and the impaired antibody production in immunized elderly were discussed. To a more detailed level, the possible effects of aerobic training on NK cell activity and the molecular mechanisms that could explain a better antiviral response in active older persons and master athletes were explored.

### **COULD LIFELONG PHYSICAL EXERCISE CREATE AN ANTI-INFLAMMATORY SCENARIO THAT ALLEVIATES THE “CYTOKINE STORM” PHENOMENON?**

Clinical studies have identified a cytokine storm (sustained responses of cytokines and chemokines) in critical patients with COVID-19 [21], pointing that patients in this condition present a higher incidence of immune disorders, acute respiratory distress syndrome, and/or extrapulmonary multiple-organ failure, which are the critical factors for COVID-19 exacerbation or even death [22]. In particular, increased levels of cytokines, including interleukin (IL)-1 $\beta$ , IL-2, IL-6, IL-8, IL-10, IL-17, interferon- $\gamma$  (IFN- $\gamma$ ), tumor necrosis factor (TNF), granulocyte colony-stimulating factor,  $\gamma$ -induced protein 10, monocyte chemoattractant protein 1, macrophage inflammatory protein (MIP)-1 $\alpha$  (MIP1- $\alpha$ ), and other molecules, characterize the cytokine storm [23–25].

Whereas the cytokine storm seems to be a key pathophysiological mechanism in elderly COVID-19 patients [23], regular endurance training may play a role in lowering some markers of systemic inflammation and regulating important metabolic and physiological muscle parameters with

aging [3,26]. In general, a single bout of exercise induces an inflammatory response similar to that caused by infection or trauma. The classical proinflammatory cytokines TNF and IL-1 $\beta$  are not normally increased after exercise [27,28], indicating that the cytokine cascade induced by exercise clearly differs from the cytokine cascade caused by infection. The augmentation in IL-6 concentrations during exercise seems to be responsible for the further increase in the concentrations of anti-inflammatory cytokines, such as IL-10 and IL-1 receptor antagonist (IL-1RA), by stimulating cortisol release and reducing TNF concentrations [29]. Besides, elevated concentrations of IL-6 from skeletal muscle trigger an anti-inflammatory signaling cascade that inhibits the secretion of proinflammatory cytokines, such as TNF or IL-1 $\beta$ , downturn the secretion of C-reactive protein from the liver [8], down-regulate the monocyte Toll-like receptor expression at both mRNA and cell surface protein levels, and lastly inhibit the I $\kappa$ B kinase/nuclear factor- $\kappa$ B (NF- $\kappa$ B) pathway [30,31]. A decreased plasma IL-6 level in response to regular exercise is illustrated as a normal training adaptation [32]. Studies have shown that acute and regular exercise can reduce the activation of this inflammatory signaling pathway (for review, see [31]) in master athletes [33].

IL-1 $\beta$  and TNF are two cytokines related to the inflammatory cascade. High levels of IL-1 $\beta$  are expected after muscle damage, whereas exercise can or cannot increase IL-1 $\beta$  levels [34]. Conversely, IL-1RA is an anti-inflammatory cytokine secreted primarily by monocytes and macrophages, and inhibits the proinflammatory actions of IL-1 $\beta$ . In particular, IL-1RA prevents inflammatory processes by blocking proinflammatory cytokine IL-1-induced signal transduction, creating an anti-inflammatory balance to the proinflammatory stimulus induced by IL-1 $\beta$  [8]. The equilibrium between IL-1 and IL-1RA in local tissues influences the potential development of inflammatory disease and its structural damage. Increased levels of IL-1RA have been observed in master athletes compared to age-matched sedentary subjects and young individuals [33].

Still, in an IL context, IL-10 has a crucial role in preventing inflammatory and autoimmune pathologies [35]. It appears to contribute to adaptive immune response down-regulation and reduction of inflammatory-induced tissue damage. The latter IL-10 contribution is mediated by decreased major histocompatibility complex molecule expression and the intercellular adhesion molecule-1 and costimulatory CD80 and CD86 molecules in antigen-presenting cells [8]. Likewise, IL-10 interferes with or completely inhibits the expression of various proinflammatory cytokines and other soluble mediators. Thus, IL-10 is a potent promoter of an anti-inflammatory state [36–38]. Exercise-induced IL-10 secretion may be associated with increased circulating regulatory T (Treg) cells, the primary source of IL-10 within the body [8]. Handzlik et al. showed that high training loads are related to higher resting Treg cell count and superior IL-10 production, resulting in antigen stimulation [39].

The authors' laboratory corroborated these findings, demonstrating adaptive responses to exercise in master athletes, which are translated to the sustained number and function of Treg cells and elevated *IL-10* gene expression [2].

Physical fitness status also promotes a positive impact in the monocyte/macrophage phenotype outcome and consequently in the anti-inflammatory response. IL-6 released during exercise drives differentiation to an anti-inflammatory state while stimulating IL-10 and IL-1RA release mediated by peroxisome proliferator-activated receptor- $\gamma$  in these cells [40]. Intracellular mechanisms in monocytes/macrophages mediating the transcriptional changes in response to anti-inflammatory cytokines may be related to energetic sensor activation, such as Adenosine monophosphate-activated protein kinase (AMPK) [41]. Acute exercise-induced immunometabolic alterations in these cells are driven by lower glucose levels, leading to AMP accumulation, which in turn stimulates AMPK activity, potentially inhibiting NF- $\kappa$ B transcription factor, and resulting in a sustained anti-inflammatory scenario [42]. This immunometabolic profile supports the known anti-inflammatory effects of physical exercise [43].

This exercise-induced cytokine response is one of several other mechanisms by which exercise exerts anti-inflammatory effects [8]. The overall anti-inflammatory profile exhibited by master athletes could likely alleviate the cytokine storm induced by COVID-19 infection, preventing health deterioration in general.

### **VACCINE, AGING, AND EXERCISE TRAINING: WHAT COULD WE EXPECT DURING COVID-19 INFECTION?**

Vaccination is the primary pharmacological strategy to reduce the incidence and severity of viral diseases. Here, how aging-associated immunosenescence could reduce vaccination efficacy on those more vulnerable to the severe form of COVID-19, the elderly population, is discussed [44]. Additionally, how physical activity could improve the vaccination response in the elderly is addressed.

Antibody production after vaccination in the elderly can be decreased. For instance, antibody titers produced after the current influenza vaccine are 70% to 90% effective in young adults but only 17% to 53% in the elderly, revealing a huge impairment in vaccination efficiency [45,46]. This immune system's impaired ability to respond to vaccines can be explained by several modifications on the antigen presentation system and the reduced immune cells' effector function. Besides a defective antigen presentation system, dendritic cells showing reduced distribution and migratory capacity have also been described during the aging process [47]. Furthermore, reduced antigen-induced phagocytosis and IFN- $\gamma$  and IL-12 secretion were observed, causing decreased lymphocyte proliferation and activation.

Of note, sirtuin-1 (SIRT-1) is essential for dendritic cells' correct mediation of immune responses. It was recently demonstrated that the specific deletion of SIRT-1 was sufficient to induce an excess of reactive oxygen species and impaired cytokine production in bone marrow dendritic cells infected with respiratory syncytial virus [48]. Disrupted dendritic cell response in SIRT-1 knockouts was restored by inhibiting acetyl-CoA carboxylase [48]. Thus, it is possible that aging-associated mitochondrial dysfunction in dendritic cells could be sufficient to disturb an immune response.

At the same time, aging also reduces naive circulating lymphocytes and CD4<sup>+</sup>/CD8<sup>+</sup> ratio [49], although, within CD8<sup>+</sup> and CD4<sup>+</sup> T memory lymphocytes, the late differentiated effector-memory subset will increase, showing higher CD45RA, CCR7<sup>+</sup>, and CD62L<sup>-</sup> expression [50]. Also, effector-memory lymphocytes show a reduction in IL-2 expression and consequently in clonal expansion and cytolytic activity [46].

Overall, besides reduced antigen presentation and CD8<sup>+</sup> T-cell cytotoxicity, diminished NK cell and B lymphocyte activity has been reported for elderly populations [49]. Aging reduces memory B-cell function and long-lived plasma cells, leading to reduced antibody production after vaccination [51].

Lifelong athletes show many adaptations at the molecular level that slow down immunosenescence. This population has presented better responses to vaccination with increased antibody production than age-matched controls [52]. Recently, Wong et al. have also observed that elderly women with increased fitness levels exhibited higher antibody titers 18 months after influenza immunization [53].

An array of immunological parameters is affected by exercise training in the elderly, explaining this improvement in vaccine response. For instance, in master athletes, the reduction of CD8<sup>+</sup> and CD4<sup>+</sup> T senescent central memory cells have been reported [1], besides exercise training inducing the chronic turnover of lymphocytes and causing increased apoptosis of senescent lymphocytes while increasing naive T-cell mobilization [54]. Furthermore, regular exercise stimulated memory B lymphocytes and antibody-producing cells function, as athletes showed high levels of circulating antibodies [55]. Wong et al. showed that high fitness older women had increased gene expression involved with phagocytic function in monocytes/macrophages accompanied by increased antibody production by B lymphocytes [53].

Regarding NK cells, the most important subsets in fine-tuning the antiviral response are the (i) CD56<sup>bright</sup>CD16<sup>-</sup> cells that produce high levels of cytokines and rapid proliferation and (ii) CD56<sup>dim</sup>CD16<sup>+</sup> cells that are a mature subset with high cytotoxic capacity [56]. In the elderly, this subset of lymphoid immune innate cells showed increased surface markers of exhausted NK cells (CD56<sup>-</sup>CD16<sup>+</sup>) that present dysfunctional cytokine production, impaired cytotoxic activity, and reduced proliferation [57]. NK cells' correct functioning is essential for the immune system to fight viral

infections. The antiviral response of NK cells is mediated by the expression of cytotoxic molecules (i.e., granzymes and perforin) or the direct interaction with death receptors [58]. Also, NK cells produce many types of cytokines and chemokines that recruit other immune cells [59]. NK-mediated immunity can be achieved in the presence or absence of antibodies [58].

As mentioned above, NK cells have a central role in antiviral response. Next, the effects of exercise on NK cells' immunometabolism regulation are described in detail. It is hypothesized that NK metabolic adaptation has a major responsibility for improving the antiviral response in master athletes.

It is noteworthy that the vulnerable population developing the severe form of COVID-19 is the same population that shows reduced vaccine efficiency. Thus, lifelong exercise training could be an efficient approach to improve not only the immune response against viral pathogens but also vaccine efficiency in the older population.

#### **EFFECTS OF LIFELONG AEROBIC TRAINING ON NK CELL ACTIVITY: DO MASTER ATHLETES HAVE A BETTER ANTIVIRAL RESPONSE?**

Two important factors, low body fat mass and healthy nutritional habits, can help explain the possible low susceptibility to viral infections in master athletes. Chiappetta et al. (2020) have reported that obese people with hyperinflammation are more susceptible to the moderate or severe form of COVID-19 infection [60]. Continuous overnutrition leads to excessive availability of nutrients in the circulation, mainly glucose and fatty acids [41]. These nutrients exert a sufficient regulation on metabolic pathways, mostly regulated by two cellular energy sensors: AMPK and mammalian target of rapamycin (mTOR) [61]. Clinical findings showed that hyperglycemia leads to poor prognosis in COVID-19 patients. In contrast, diabetic patients treated with metformin (a pharmacological activator of AMPK) showed a reduction in mortality [62]. The activation and inhibition of these pathways can change cell function, especially immune cells, such as NK cells.

The main functions of activated NK cells are (i) killing virus-infected or malignantly transformed cells and (ii) triggering the adaptive immune response through the release of cytokines (e.g., IFN- $\gamma$  and TNF) and chemokines (e.g., MIP-1 $\beta$ ). When encountering target cells, NK cells release the content of their cytotoxic granules (e.g., perforin and granzymes) into the immune synapse to kill their targets. In the last years, the scientific literature has highlighted the importance of NK cell metabolism in facilitating robust NK cell effector functions [63].

Resting human NK cells have low basal metabolic rates of glycolysis and oxidative phosphorylation to meet their homeostatic needs. When NK cells become activated, they change their metabolism. After overnight stimulation with IL-2 or IL-12 plus IL-15, activated NK cells undergo dramatic metabolic reprogramming, up-regulating rates of glucose uptake

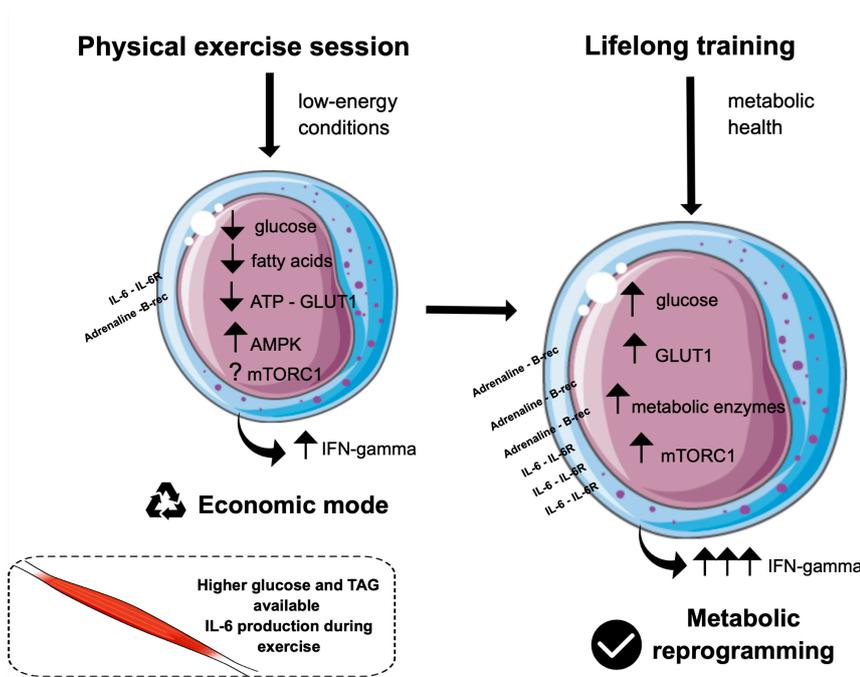
and glycolysis [64], the mTORC1 activity being essential for attaining this elevated glycolytic state [65]. When NK cells are stimulated over longer periods, IL-2 drives mTORC1-dependent glycolytic reprogramming of NK cells, sustaining NK cell effector functions and allowing them to work in parallel with the adaptive immune response [66]. mTORC1 inhibition by rapamycin specifically diminished IL-2 and IL-12 stimulation-dependent increase of glycolysis, leading to impaired cell growth and limited IFN- $\gamma$  production and granzyme B expression [65]. Interestingly, in a lipid-rich environment, such as in obesity, the inhibition of mTORC1 activation in human NK cells inhibits their effector function [67]. Thus, the mTORC1 pathway is crucial for NK cell function [67].

Notably, strategies that reduce metabolic rates in mouse NK cells inhibit cytokine-induced proliferation and impair NK cell cytotoxicity. Murine cytomegalovirus-infected mice treated with the glycolytic inhibitor 2-deoxy-D-glucose presented impaired clearance of NK-specific target cells and increased viral burden [68]. Under conditions of glucose limitation, a plasticity of the immune cell metabolic pathways occurs; however, it is still poorly understood how NK cell metabolic pathways adapt to metabolically restrictive environments.

Altogether, a metabolic reprogramming of NK cells during an exercise session and more robustly with lifelong exercise training is proposed. During a physical exercise session, NK cells experience low-energy conditions. This could acutely favor the activation of energetic sensors (here, the AMPK is suggested), mainly stimulating GLUT1 translocation to the cell membrane for glucose uptake purposes. This momentarily active, insufficient condition during exercise drives NK cells to function in an “economical mode.” However, after the end of the exercise training session, with adequate rest and nutrition, the stimulation of the mTOR pathway begins, in turn promoting metabolic reprogramming in an attempt to create a more efficient cellular and functional profile. Thus, exercise-induced activation of mTORC1 via muscle-derived IL-6 and adrenaline secretion promotes an efficient NK cell pathogen elimination. This repeated stimulus, given the high volume and intensity of exercise training sessions, enhances NK cell adaptations over time. Therefore, lifelong exercise would induce a metabolic remodeling in NK cells: increasing glycogen (glucose) availability, facilitating GLUT1 translocation, and enhancing the expression of glycolytic pathway metabolic enzymes. This hypothesis is illustrated in Figure 1.

Lifelong exercise training could delay age-related decrements in immunity. Lifelong exercisers would display a preserved immune response to exercise, indicating that they are better prepared to adapt (or respond) to new immune challenges, such as a viral infection. Metabolic programming in immune cells could be a determinant for better antiviral response, such as that needed to fight SARS-CoV-2. Studies on this topic could shed light on the understanding of how lifelong exercise could lead to immunoprotection.

In summary, repetitive episodes of exercise throughout life may induce metabolic changes (systemic and intracellular), forcing the immune cells to work in different metabolic scenarios (catabolic and anabolic conditions) and promoting important intracellular adaptations for a better response against viral infections. This new metabolic and anti-inflammatory profile could help block/prevent the cytokine storms observed during COVID-19 infection. The fundamental message is that people should expose immune cells to energy deprivation situations during the day, for instance, by adopting regular physical exercise during their lifetime. This “simple” habit can promote greater immune system efficiency.



**Figure 1.** Immunometabolism of NK cells during an exercise session and with lifelong exercise training.

**AUTHOR CONTRIBUTIONS**

FSL and JCR conceived the present idea. LGM, FSL and JCL wrote the paper with input from all authors. AMT and RVTS did a critical revision of the article. All authors read and approved the final manuscript.

**CONFLICTS OF INTEREST**

The authors declare that there is no conflict of interest.

**FUNDING**

The present manuscript was accomplished with support from CAPES, Coordination for the Improvement of Higher Education Personnel, Brazil (Capes-print Unesp, Capes 001) and São Paulo Research Foundation (Fapesp 2019/10187-7).

## ACKNOWLEDGMENTS

The present study was conducted in honor of the 15-year anniversary of the death of Professor Luis Fernando Pereira Bicudo Costa Rosa, an eminent exercise immunology researcher in Brazil, whose ideas, erudition, critical sense and honesty continue to inspire us.

## REFERENCES

1. Minuzzi LG, Rama L, Chupel MU, Rosado F, Dos Santos JV, Simpson R, et al. Effects of lifelong training on senescence and mobilization of T lymphocytes in response to acute exercise. *Exerc Immunol Rev.* 2018;24:72-84.
2. Minuzzi LG, Rama L, Bishop NC, Rosado F, Martinho A, Paiva A, et al. Lifelong training improves anti-inflammatory environment and maintains the number of regulatory T cells in masters athletes. *Eur J Appl Physiol.* 2017;117:1131-40.
3. Mikkelsen UR, Couppé C, Karlsen A, Grosset JF, Schjerling P, Mackey AL, et al. Life-long endurance exercise in humans: circulating levels of inflammatory markers and leg muscle size. *Mech Ageing Dev.* 2013;134:531-40.
4. Campbell JP, Turner, JE. Debunking the myth of exercise-induced immune suppression: Redefining the impact of exercise on immunological health across the lifespan. *Front Immunol.* 2018;9:1-21.
5. Sellami M, Gasmi M, Denham J, Hayes LD, Stratton D, Padulo J, et al. Effects of acute and chronic exercise on immunological parameters in the elderly aged: Can physical activity counteract the effects of aging? *Front Immunol.* 2018;9:1-17.
6. Aguiar SS, Sousa CV, Deus LA, Rosa TS, Sales MM, Neves RVP, et al. Oxidative stress, inflammatory cytokines and body composition of master athletes: The interplay. *Exp Gerontol.* 2020;130:110806.
7. Antunes BM, Campos EZ, Dos Santos RVT, Rosa-Neto JS, Franchini E, Bishop NC, et al. Anti-inflammatory response to acute exercise is related with intensity and physical fitness. *J Cell Biochem.* 2019;120:5333-42.
8. Gleeson M, Bishop NC, Stensel DJ, Lindley MR, Mastana SS, Nimmo MA. The anti-inflammatory effects of exercise: mechanisms and implications for the prevention and treatment of disease. *Nat Rev Immunol.* 2011;11:607-15.
9. Woods JA, Vieira VJ, Keylock KT. Exercise, Inflammation, and Innate Immunity. *Neurol Clin.* 2006;24:585-99.
10. Walsh NP, Gleeson M, Pyne DB, Nieman DC, Dhabhar FS, Shephard RJ, et al. Position statement. Part two: Maintaining immune health. *Exerc Immunol Rev.* 2011;17:64-103.
11. Krüger K, Mooren FC, Pilat C. The Immunomodulatory Effects of Physical Activity. *Curr Pharm Des.* 2016;22:3730-48.
12. Bigley AB, Rezvani K, Chew C, Sekine T, Pistillo M, Crucian B, et al. Acute exercise preferentially redeploys NK-cells with a highly-differentiated phenotype and augments cytotoxicity against lymphoma and multiple myeloma target cells. *Brain Behav Immun.* 2014;39:160-71.
13. Evans ES, Hackney AC, McMurray RG, Randell SH, Muss HB, Deal AM, et al.

- Impact of Acute Intermittent Exercise on Natural Killer Cells in Breast Cancer Survivors. *Integr Cancer Ther.* 2015;14:436-45.
14. Idorn M, Hojman P. Exercise-Dependent Regulation of NK Cells in Cancer Protection. *Trends Mol Med.* 2016;22:565-577.
  15. Yuki K, Fujiogi M, Koutsogiannaki S. COVID-19 pathophysiology: A review. *Clin Immunol.* 2020;215:108427.
  16. Mäkinen TM, Juvonen R, Jokelainen J, Harju TH, Peitso A, Bloigu A, et al. Cold temperature and low humidity are associated with increased occurrence of respiratory tract infections. *Respir Med.* 2009;103:456-62.
  17. Foxman EF, Storer JA, Fitzgerald ME, Wasik BR, Hou L, Zhao H, et al. Temperature-dependent innate defense against the common cold virus limits viral replication at warm temperature in mouse airway cells. *Proc Natl Acad Sci.* 2015;112:827-32.
  18. Zhao W, An H, Zhou J, Xu H, Yu Y, Cao X. Hyperthermia differentially regulates TLR4 and TLR2-mediated innate immune response. *Immunol Lett.* 2007;108:137-42.
  19. Saghazadeh A, Rezaei N. Immune-epidemiological parameters of the novel coronavirus—a perspective. *Expert Rev Clin Immunol.* 2020;16:465-70.
  20. Wang F, Nie J, Wang H, Zhao Q, Xiong Y, Deng L, et al. Characteristics of Peripheral Lymphocyte Subset Alteration in COVID-19 Pneumonia. *J Infect Dis.* 2020;221(11):1762-9. doi: 10.1093/infdis/jiaa150
  21. Channappanavar R, Perlman S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. *Semin Immunopathol.* 2017;39:529-39.
  22. Ye Q, Wang B, Mao J. The pathogenesis and treatment of the ‘Cytokine Storm’ in COVID-19. *J Infect.* 2020;80:607-13.
  23. Meftahi GH, Jangravi Z, Sahraei H, Bahari Z. The possible pathophysiology mechanism of cytokine storm in elderly adults with COVID-19 infection: the contribution of “inflamm-aging”. *Inflamm Res.* 2020;69:825-39.
  24. Li H, Liu L, Zhang D, Xu J, Dai H, Tang N, et al. SARS-CoV-2 and viral sepsis: observations and hypotheses. *Lancet.* 2020;395:1517-20.
  25. Cao X. COVID-19: immunopathology and its implications for therapy. *Nat Rev Immunol.* 2020;20:269-70.
  26. Nieman DC, Henson DA, Smith LL, Utter AC, Vinci DM, Davis JM, et al. Cytokine changes after a marathon race. *J Appl Physiol.* 2001;91:109-14.
  27. Reihmane D, Jurka A, Tretjakovs P, Dela F. Increase in IL-6, TNF- $\alpha$ , and MMP-9, but not sICAM-1, concentrations depends on exercise duration. *Eur J Appl Physiol.* 2013;113:851-8.
  28. Steensberg A, Fischer CP, Keller C, Møller K, Pedersen BK. IL-6 enhances plasma IL-1ra, IL-10, and cortisol in humans. *Am J Physiol Endocrinol Metab.* 2003;285:E433-7.
  29. Sallam N, Laher I. Exercise Modulates Oxidative Stress and Inflammation in Aging and Cardiovascular Diseases. *Oxid Med Cell Longev.* 2016;2016:7239639.
  30. Rodriguez-Miguel P, Fernandez-Gonzalo R, Almar M, Mejías Y, Rivas A, de Paz JA, et al. Role of Toll-like receptor 2 and 4 signaling pathways on the inflammatory response to resistance training in elderly subjects. *Age.*

- 2014;36:9734.
31. Röhling M, Herder C, Stemper T, Müssig, K. Influence of Acute and Chronic Exercise on Glucose Uptake. *J Diabetes Res.* 2016;2016:1-33.
  32. Fischer CP. Interleukin-6 in acute exercise and training: what is the biological relevance? *Exerc Immunol Rev.* 2006;12:6-3.
  33. Minuzzi LG, Chupel MU, Rama L, Rosado F, Muñoz VR, Gaspar RC, et al. Lifelong exercise practice and immunosenescence: Master athletes cytokine response to acute exercise. *Cytokine.* 2019;115:1-7.
  34. Gagnon DD, Gagnon SS, Rintamäki H, Törmäkangas T, Puukka K, Herzig K-H, et al. The Effects of Cold Exposure on Leukocytes, Hormones and Cytokines during Acute Exercise in Humans. *PLoS One.* 2014;9:e110774.
  35. Fujio K, Okamura T, Yamamoto K. The Family of IL-10-secreting CD4<sup>+</sup> T cells. *Adv Immunol.* 2010;105:99-130.
  36. Caton AJ, Weissler KA. Regulatory cells in health and disease. *Immunol Rev.* 2014;259:5-10.
  37. Speaker KJ, Cox SS, Paton MM, Serebrakian A, Maslanik T, Greenwood BN, et al. Six weeks of voluntary wheel running modulates inflammatory protein (MCP-1, IL-6, and IL-10) and DAMP (Hsp72) responses to acute stress in white adipose tissue of lean rats. *Brain Behav Immun.* 2014;39:87-98.
  38. Saraiva M, O'Garra A. The regulation of IL-10 production by immune cells. *Nat Rev Immunol.* 2010;10:170-81.
  39. Handzlik MK., Shaw AJ, Dungey M, Bishop NC, Gleeson M. The influence of exercise training status on antigen-stimulated IL-10 production in whole blood culture and numbers of circulating regulatory T cells. *Eur J Appl Physiol.* 2013;113:1839-48.
  40. Silveira LS, Antunes BMM, Minari ALA, Dos Santos RVT, Neto JCR, Lira FS. Macrophage Polarization: Implications on Metabolic Diseases and the Role of Exercise. *Crit Rev Eukaryot Gene Expr.* 2016;26:115-32.
  41. Von Ah Morano AE, Dorneles GP, Peres A, Lira FS. The role of glucose homeostasis on immune function in response to exercise: The impact of low or higher energetic conditions. *J Cell Physiol.* 2020;235(4):3169-88. doi: 10.1002/jcp.29228
  42. Salminen A, Hyttinen JMT, Kaarniranta K. AMP-activated protein kinase inhibits NF-κB signaling and inflammation: impact on healthspan and lifespan. *J Mol Med.* 2011;89:667-76.
  43. de Souza Teixeira AA, Lira FS, Pimentel GD, de Souza CO, Batatinha H, Biondo LA, et al. Aerobic exercise modulates the free fatty acids and inflammatory response during obesity and cancer cachexia. *Crit Rev Eukaryot Gene Expr.* 2016;26(3):187-98.
  44. Berg-Weger M, Schroepfer T. COVID-19 Pandemic: Workforce Implications for Gerontological Social Work. *J Gerontol Soc Work.* 2020;1-6. doi: 10.1080/01634372.2020.1772934
  45. Goodwin K, Viboud C, Simonsen L. Antibody response to influenza vaccination in the elderly: A quantitative review. *Vaccine.* 2006;24(8):1159-69.
  46. Simpson RJ, Lowder TW, Spielmann G, Bigley AB, LaVoy EC, Kunz H. Exercise and the aging immune system. *Ageing Res Rev.* 11, 404-20.

47. Panda A, Arjona A, Sapey E, Bai F, Fikrig E, Montgomery RR, et al. Human innate immunosenescence: causes and consequences for immunity in old age. *Trends Immunol.* 2009;30(7):325-33. doi: 10.1016/j.it.2009.05.004
48. Elesela S, Morris SB, Narayanan S, Kumar S, Lombard DB, Lukacs NW. Sirtuin 1 regulates mitochondrial function and immune homeostasis in respiratory syncytial virus infected dendritic cells. *PLoS Pathog.* 2020;16:e1008319.
49. Duggal NA, Niemiro G, Harridge SDR, Simpson RJ, Lord JM. Can physical activity ameliorate immunosenescence and thereby reduce age-related multi-morbidity? *Nat Rev Immunol.* 2019;19:563-72.
50. Henson SM, Riddell NE, Akbar AN. Properties of end-stage human T cells defined by CD45RA re-expression. *Curr Opin Immunol.* 2012;24:476-81.
51. Nikolich-Zugich J. The twilight of immunity: Emerging concepts in aging of the immune system review-article. *Nat Immunol.* 2018;19:10-19.
52. de Araújo, AL, Silva LCR, Fernandes JR, Matias MST, Boas LS, Machado CM, et al. Elderly men with moderate and intense training lifestyle present sustained higher antibody responses to influenza vaccine. *Age.* 2015;37:105.
53. Wong GCL, Narang V, Lu Y, Camous X, Nyunt MSZ, Carre C, et al. Hallmarks of improved immunological responses in the vaccination of more physically active elderly females. *Exerc Immunol Rev.* 2019;25:20-33.
54. Krüger K, Alack K, Ringseis R, Mink L, Pfeifer E, Schinle M, et al. Apoptosis of T-Cell Subsets after Acute High-Intensity Interval Exercise. *Med Sci Sports Exerc.* 2016;48,2021-9.
55. Nieman DC, Nehlsen-Cannarella SL. The Effects of Acute and Chronic Exercise on Immunoglobulins. *Sports Medicine.* 1991;11(3):183-201. doi: 10.2165/00007256-1991111030-00003
56. Freud AG, Mundy-Bosse BL, Yu J, Caligiuri MA. The Broad Spectrum of Human Natural Killer Cell Diversity. *Immunity.* 2017;47:820-33.
57. Tarazona R, Sanchez-Correa B, Casas-Avilés I, Campos C, Pera A, Morgado S, et al. Immunosenescence: limitations of natural killer cell-based cancer immunotherapy. *Cancer Immunol Immunother.* 2017;66:233-45.
58. Fiscaro P, Rossi M, Vecchi A, Acerbi G, Barili V, Laccabue D, et al. The Good and the Bad of Natural Killer Cells in Virus Control: Perspective for Anti-HBV Therapy. *Int J Mol Sci.* 2019;20(20):5080.
59. Vivier E, Raulet DH, Moretta A, Caligiuri MA, Zitvogel L, Lanier LL, et al. Innate or adaptive immunity? The example of natural killer cells. *Science.* 2011;331:44-9.
60. Chiappetta S, Sharma AM, Bottino V, Stier C. COVID-19 and the role of chronic inflammation in patients with obesity. *Int J Obes.* 2020;44:1790-2.
61. Batatinha HAP, Biondo LA, Lira FS, Castell LM, Rosa-Neto JC. Nutrients, immune system, and exercise: Where will it take us? *Nutrition.* 2019;61:151-6.
62. Luo P, Qiu L, Liu Y, Liu X-L, Zheng J-L, Xue H-Y, et al. Metformin Treatment Was Associated with Decreased Mortality in COVID-19 Patients with Diabetes in a Retrospective Analysis. *Am J Trop Med Hyg.* 2020;103(1):69-72. doi: 10.4269/ajtmh.20-0375
63. O'Brien KL, Finlay DK. Immunometabolism and natural killer cell responses. *Nat Rev Immunol.* 2019;19:282-90.

64. Keating SE, Zaiatz-Bittencourt V, Loftus RM, Keane C, Brennan K, Finlay DK, et al. Metabolic Reprogramming Supports IFN- $\gamma$  Production by CD56 bright NK Cells. *J Immunol*. 2016;196(6):2552-60. doi: 10.4049/jimmunol.1501783
65. Donnelly RP, Loftus RM, Keating SE, Liou KT, Biron CA, Gardiner CM et al. mTORC1-Dependent Metabolic Reprogramming Is a Prerequisite for NK Cell Effector Function. *J. Immunol*. 2014;193(9):4477-84. doi: 10.4049/jimmunol.1401558
66. Gardiner CM, Finlay DK. What fuels natural killers? Metabolism and NK cell responses. *Front Immunol*. 2017;8:367.
67. Michelet X, Dyck L, Hogan A, Loftus RM, Duquette D, Wei K, et al. Metabolic reprogramming of natural killer cells in obesity limits antitumor responses. *Nat Immunol*. 2018;19:1330-40.
68. Mah AY, Rashidi A, Keppel MP, Saucier N, Moore EK, Alinger JB, et al. Glycolytic requirement for NK cell cytotoxicity and cytomegalovirus control. *JCI Insight*. 2017;2(23):e95128. doi: 10.1172/jci.insight.95128

How to cite this article:

Minuzzi LG, Teixeira AM, Thomatieli-Santos RV, Rosa-Neto JC, Lira FS. Immunometabolism and Covid-19: Could Lifelong Exercise Training Have a Protective Effect? *Immunometabolism*. 2021;3(1):e210001. <https://doi.org/10.20900/immunometab20210001>